Molecular study of bacteria Pantoea spp. in Diwaniyah city, Iraq

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Abstract

Pantoea spp. possesses many virulent genes that help to cause many human infections. Therefore, the present study was comprehensive of clinical and environmental samples to determine the extent of the spread of *Pantoea* spp. in the hospitals in the city of Diwaniyah.

A total of 623 samples were collected and distributed as483 samples of different clinical cases (77.52%) and 140 environmental samples (22.47%) from Diwaniyah city hospitals. The results showed that 24 isolates of *Pantoea* spp.s had been confirmed using API-20E Polymerase chain reaction technology.

All isolates of *Pantoea* spp were tested for the prevalence of virulence genes (*rcsB*, *hpaA*, , *hrc*, *rcsA*), using the polymerase chain reaction (PCR) technique. The highest incidence of *hrc* gene (70.83) .The presence of *rcsA* gene was 68.42% with 13 isolates. The current study did not record any presence of the virulence genes(*rcsB*, *hpaA*) among the isolates of *Pantoea* spp.

Introduction

Pantoea spp. bacteria is known to belong to the enterobacteriaceae family, which is negative on Gram Stain (1).

The main problem is that the *Pantoea* spp. bacteria are associated with opportunistic diseases, although they are not a compulsory infection (2).

Pantoea spp. have *hrc*, *rcsA* genes that are responsible for the production of exopolysaccharide (EPS) were identified which help with the process of bacterial adhesion and the enzymes that have a role in the pathogenesis of these bacteria, such as Cellulase enzyme, which helps analyze the walls of plant cells consisting of a thick layer of cellulose polymer that provides protection of the plant from the penetration of microbes and increasing the ferocity of the bacteria causing the disease inside host cells (3).

rcsA and *rcsB* genes are regulatory genes that control the production of capsule polysaccharide synthesis (EPS) (exopolysaccharide) (4).

The *hpaA* gene also increases the secretion of pilius and confuses host defense devices and increases virulence in them because pilius increases the susceptibility of bacteria to adhering to host tissue and thereby manipulating cellular host activity (5).

Collection of Samples

As many as 483 clinical and 140 environmental samples were collected from Diwaniyah hospitals including Diwaniyah General Teaching Hospital, Women's and Children Teaching Hospital Children, Burns Specialist and Consultancy Center, and the Public Health Laboratory for the period from October 2017 to June 2018. Swabs were collected from different clinical samples, which included 82 samples and swaps from pharynx (23), ears (49), dialysis (83), stool (solid and diarrhea) (27), pus (31) (27), wounds (44), burns (70) and cough (74). Environmental samples include floor (30), medical tools (40), equipment (20), walls (30) and beds (20). Medical cotton swabs containing the center of the transport media swabs were used in sampling to ensure the vitality of the isolates, and then the necessary tests in vitro and biochemical laboratory of the Faculty of Science / University of Al-Qadisiyah for the bacteria *Pantoea* spp. were applied.

DNA primers

Gene 16rsRNA primers were used for the diagnosis of *Pantoea* spp. The primer genes responsible of identifying virulence genes in these bacteria were used. All primers were designed using NCBI-Genbank and primer3plus design. These primers were prepared by Bioneer Co. in Korea (Table 1).

Primer	Sequence		Amplicon	Gene bank code
16srRNA	F	CCTGGACAAAGACTGACGCT		
	R	CGCTTCTCTTTGTATGCGCC	523bp	FR832419.1
	R	TTCCATGATGCCGCTCACAT		
rcsA	F	AAGTCCATCCGTTGACGCTT	395bp	
	R	CAATTTACCGATGGCTGCCG	0,000	M60621.1
rcsB	F	GAACAACAACCCGGCCATTC	128hn	
	R	CTTTTGATAAGGCCGCCACG	438bp	Y09848.1
Hrc	F	TACAGCCAGCTGATTGCCAA		U56662.2:35
			647bp	05-5535
	R	CGCTGTGAGATTTCATGGCG		

Table 1: DNA primers used in this study

	R	AGATGGGCTGTGCGTCATC		
	R	CAACACGATGCAAGAGACGC		
	F	TGGGCAGTAACGATGTGCAT		NC-
			519bp	003197.2:11
HpaA		AAAGTTTCAGTTCACCGCGC	-	05402
	R			95402-
				1196306

Forword, R: Reverse F

DNA-Amplifying PCR Thermocycler Programs

Polymerization enzyme chain reaction was applied using PCR Thermocycler as in the following table.

Gene	Temperature (°C) / Time					Cycles
Name	Initial	Final	Numbr			
	Denaturat-	Denaturation	Annealing	Extension	Extension	
	ion / time				/ time	
16srRNA						
hrc						
rcsA	95/5 min	95/30Sec.	58/30Sec	72/1min	72 /5min	35
rcsB						
	95/5 min	95 /30Sec.	59/1 min	72/1 min	72/5min	35

Table 2: temperature conditions used in the PCR

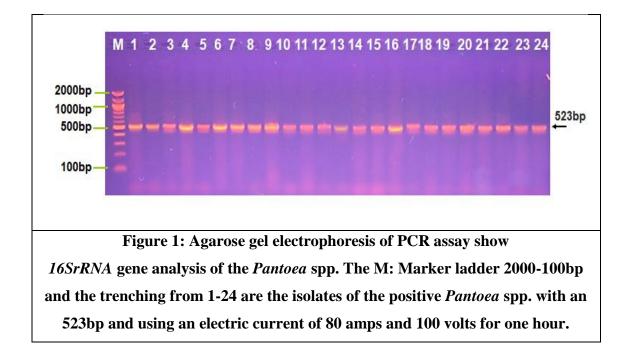
hpaA	95/5min	95/30sec.	57/30sec	72/3min	72/5min	35

Results and Discussions

Diagnosis of Pantoea spp.using PCR technology

The isolates of *Pantoea* spp. in this study, have been diagnosed using the technique of polymerase chain reaction (Monoplex PCR) with 16S rRNA gene. The results in Figure 1 show that all isolates contain 16S rRNA gene with molecular weight (523 bp), which represents the diagnostic gene for these bacteria. This confirms that the isolates belong to Pantoea spp. genes. The PCR technique was 100% sensitive to the diagnosis of isolates during this study.

The study of (6) indicated that 16S rRNA gene was used to detect *Pantoea* spp. in 18 isolates confirming that 9 isolates belonged to this bacterial family. However, the study of (7) diagnosed *Pantoea* spp. isolates that are pathogenic to humans and animals using a 16S rRNA gene with 95.6% of its isolates containing this gene.



Virulence Genes in *Pantoea* spp. Isolates

The detection of the presence of virulence genes in the isolates of *Pantoea* spp. is of great importance in epidemiology in Iraq in general because of the lack of local studies on the prevalence of virulent genes in the clinical isolates of the bacteria *Pantoea* spp., and in the hospitals of the city of Diwaniyah in particular since there have been no previous studies of the presence of isolates of these bacteria in hospitals with the spread of genes virulence in In this study.

Table 3 shows the distribution and spread of four genes (*hrc, rcsA, rcsB, hpaA*) in the isolates of *Pantoea* spp.

The results of the current study showed the presence of the *hrc* gene in 17 isolates of *Pantoea* spp. (%70.83). The number of *P. agglomerans* isolates containing this gene was 8 (33.33%) and as 9 for *P.calida* (37.5%) (Figure2). This result is consistent with those of (8) about the spread of this gene in *Pantoea* spp. isolates, especially in *P.agglomerans*

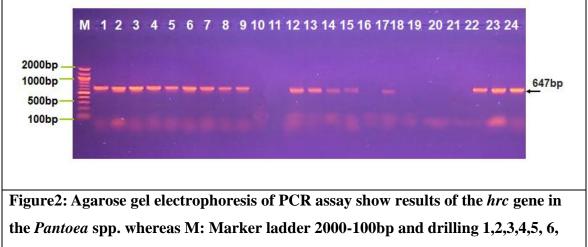
and *P.calida*. also, (9) recorded the presence of *hrc* in high quantities in *P. agglomerans* compared to other bacterial species belonging to the intestinal family.

As for the presence of *rcsA* gene in the isolates of the studied *Pantoea* spp., 68.42% (13 isolates) was recorded figure(3), *P. agglomerans* and *P.calida* recorded 25% and 29.16% (6,7 isolates)respectively. This is what was noted by (10) as the presence of *rcsA* gene in *Pantoea* spp. which is encoded to produce EPS as one of the factors of virulence in the pathogenic bacteria, which has a strong effect on the emergence of symptoms of the disease on the host as RcsA proteins encodes the gene to manufacture the capsule polysaccharides being important for the disease of bacteria. Sudies have indicated that the presence of gene *rcsA* and *rcsB* is crucial to the strains coexisting with the host to be converted to pathogenic strains and enhancing the formation of biofilm which also increases the virulence of pathogenic bacteria (11).

The current study did not record any presence of virulence genes (rcsB, hpaA,)figure (4,5) respectively in *Pantoea* spp. isolates (Table3). However. Also, (9) concluded that *P.agglomerans* bacteria contain hpaA gene. Studies have shown that the hpaA gene destroys host cells and immune devices and inhibits the host cells, which is contributing to increased virulence and disease progression (5). In other studies, the hpaA gene is affected by mutations, especially pathogenic bacteria, but it retains the stimulation of other pathogen-related genes. Also, studies pointed to the importance of HpaA proteins in the interference with the workings of host cells as these proteins are transferred to the host cell through the pathway in the period before the causing of severe disease symptoms (12).

Isolate #	<i>P.calida</i> containing	Isolate #	<i>P</i> .agglomer ans	Pantoea spp.	gene
	the gene		containing the gene	containing the gene	
1,4,5,6,7,9,14	(%37.5)9	2,3,8,12	(%33.33) 8	(%70.83)17	
,15,17		,13,22,2 3,24			hrc
4,6,7,9,14,15,	(%29.16) 7	10,11,1 2,19,22,	(%25)6	(%68.42)13	rcsA
		24			
-	(0%) 0	-	(0%) 0	(0%) 0	rcsB
-	(0%) 0	-	(0%) 0	(0%) 0	hpaA

Table 3: Virulence Genes Distribution (%) in Pantoea spp. Isolates



the *Pantoea* spp. whereas M: Marker ladder 2000-100bp and drilling 1,2,3,4,5, 6, 7,8,9, 12,13,14,15,17, 22,23,24), isolates isolates Germ of *Pantoea* spp. Positive for the gene with a length of 647bp. Using an 80-ampere electric current and 100

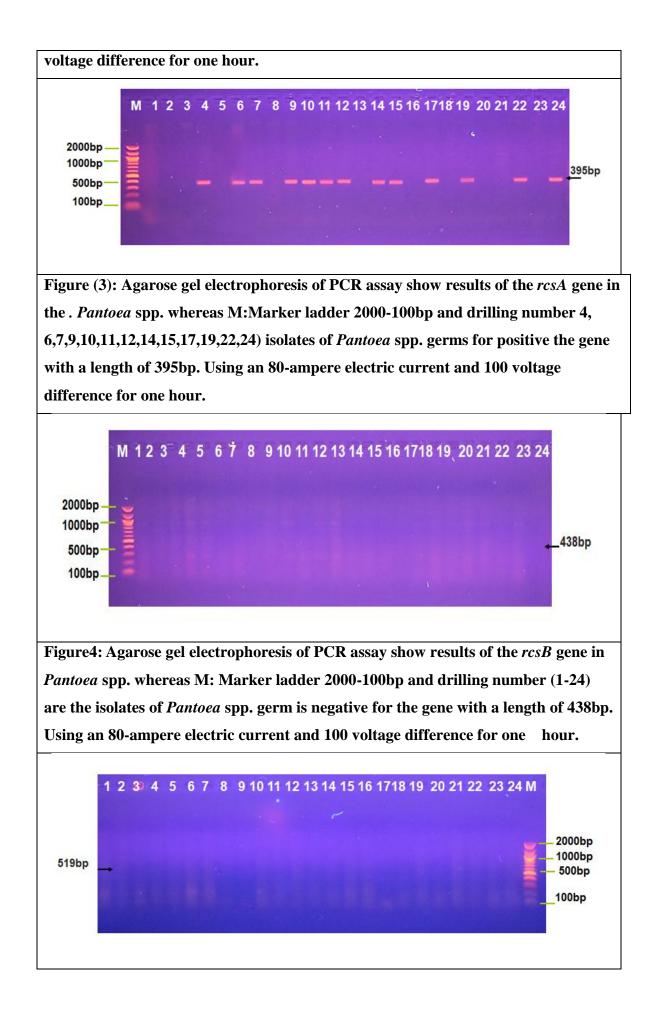


Figure5; Agarose gel electrophoresis of PCR assay show results of the *hpaA* gene in *Pantoea* spp. whereas M: Marker ladder 2000-100bp and drilling number (1-24) are the isolates of *Pantoea* spp germ . is negative for the gene with a length of 519bp. Using an 80-ampere electric current and 100 volt difference for one hour.

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