MOLECULAR CYTOGENETIC TESTING OF CHRYSOMYA BAZZIANA AND COCHLIOMYIA HOMINIVORAX (Diptera: Calliphoridae) IN AL-QADISSIYAH PROVINCE

ENTOMOLOGY Chapter-I JANUARY/Vol-9.0/Issue-I



ISSN CODE: 2456-1045 (Online) (ICV-ENT/Impact Value): 3.08 (GIF) Impact Factor: 2.174 Copyright@IJF 2017

Journal Code: ARJMD/ENT/V-9.0/I-1/C-1/JAN-2017

Website: www.journalresearchijf.com

Received: 23.12.2016 Accepted: 26.12.2016

Date of Publication: 01.02.2017

Page: 01-03



Name of the Authors:

Hussein R. Mahmood

College of sciences/university of AL-Qadisiya (IRAQ) hussein.mahmoud85@yahoo.com

Citation of the Article

Mahmood H.R (2017, January Molecular cytogenetic testing of Chrysomya bazziana and Cochliomyia hominivorax(Diptera:calliphoridae) in AL-qadissiyah province Advance Research Journal of Multidisciplinary Discoveries. , Vol. 9.0, C1, PP. 01-03 ISSN-2456-1045. from http://www.journalresearchijf.com

ABSTRACT

Chrysomya bazziana and hominivorax have 12 chromosomes, one pair is sex chromosomes and the rest are metacenteric. The karyotype morphology has no substantial differences in both species, except for the X chromosome which is subtelocentric in Chrysomya bazziana ,as metacentric in C. hominivorax and approximately, 1.3 times longer in Cochliomyia. Hominivorax. The presence of a C-band in the pericentromeric region was exploited to characterize autosomes. The sex chromosomes of both species were heterochromatic, with exception the final region of long arm of X chromosome. The detection of ribosomal genes was achieved by Molecular cytogenetic testing fluorescent In situ hybridization technique(FISH) .The position of NOR was on the sex chromosomes.

Key words:

Chrysomya - karyotype - NOR - heterochromatin - blow flies

INTRODUCTION

Chrysomya bazziana and Cochliomyia hominivorax have hygienically and veterinary importance seeing that they constitute a risk on public health and economy. They in charge of myiasis, where they lay eggs in moist skin, orifices and existing wounds (1,2). The worldwide spread and their obligation parasitism are the major reason that increases their risk. (3)

The cytogenetic variations of the natural populations of medical and economic insects play an important and decisive role the dexpansion of insecticide durability ,the implementation of genetic control as well as the former cytolgenetic studies pointed out that chromosome morphology is helpful in taxonomy (4). The heterochromatins allotment the NORs position and the obscurity or entity of sex chromosomes of various species of Muscoidea was examined, where the karyotype number 2n=12 of this group, even though the description of species that they have 2n=was demostrated, the sex pair in the latter cases was abscent (5,6,7) There was somatic pairing intimation through prophase to metaphase in most diptera and cell types studied (8). The present investigation involves the examination of the karyotype of Chrysomya bazziana and Cochliomvia hominivorax as well as the molecular cytogenetic testing (in situ hybridization) was exploited to identify the location of the nucleolar organizer regions.

MATERIALS AND METHODS

Fly rearing

Chrysomya bazziana and Cochliomyia hominivorax were collected from locations of cattle and sheep in the south of Iraq(AL-Qadissiya province). A tenuous external morphological characteristic was used to identify species. The adults were kept in nylon cages (25x25x43)cm at 25±2°C and 45-55% RH. The adults had access to sugar cane 24 h/ day and to ground beef a few hours/day. Water was always available.

Chromosome preparations

The neural ganglia of L3 larvae were exploited to get mitotic chromosomes, where 0.01 ml of colchicine was used to inject larva followed by vivisection after 45 min treatment. The dissection of brains was accomplished by fine forceps, Then, dispersal in KCl(3 ml) at 25 °C for quarter of an hour, centrifugation at 600 rpm for 10 min, and fixation in methanol. Air-drying technique was followed to prepare slides (8).The brain of larva that do not treat by colchicine was used for cytogenetic studies.

Chromosome morphology

10% Giemsa was used to stain slides for shape studies. The method remembered by **(9)** was followed for the description of the chromosomes morphology.

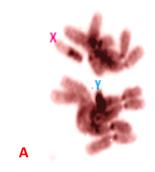
Fluorescent In situ hybridization

The baking of slides took 2 h at 80°C and 80% and 100%ethanol was used to dehydrate the baked slides. 25 mM of NaOH was used to denature chromosome for 60 sec, then washed for 10 sec. in 0.4 x SSC, 0.1% Tween, followed by dehydration by a series of 70% (precooled at -20°C), 80% and 100%ethanol for 2 min each. The composition of hybridization solution is 20 ng DIGlabelled DNA probe, 50%formamide (deionized) and 35% master mix (1 ml 20 x SSC, 1 ml dextran sulfate, 1 ml aqua

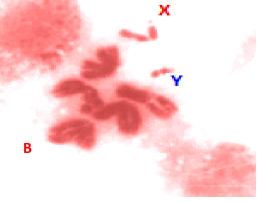
aqua bidest. and0.5 ml salmon sperm DNA (10 mg/ml, sheared to 200 - 500 bp). 10 uJ hybridization mixture was denatured at 80°C for 8 min, cooled on ice and then applied to the slide, covered with a cover-slip and sealed with rubber cement. Hybridization came by at 37°C in a humid chamber for 12 - 18 h. After two washes in 0,4 x SSC, 0.1% Tween at room temperature, anti-DIG-antibodies (fluorescein-conjugated)were applied based on the instructions of the supplier (Boehringer). Antifade (0.233 g DABCO in 0.9 ml glycerol, 0,1 ml 0.2 M Tris pH 8) containing 0.2 ug/^il propidiumiodide was put on the slide. Photos from fluorescence microscopy were taken with Kodak Ecta Goldll (400 ASA) films.

RESULTS AND DISCUSSION

The chromosomes were metacentricm in both species as illustrated in Table1, with exception for X chromosome of Chrysomya bazziana ,where was subtelocentric. karyotype length showed no major difference (37.8) µm. The X chromosome of Cochliomyia hominivorax was the longest ,while Y chromosome was too short incomparison with X chromosome for the both species. In Chrysomya bazziana, the long arm of the X chromosome has a constriction as show in figure1(A,B).In most Chrysomya species that studied, the length of sex chromosome is medium, heterochromatic, and Male factor of Y chromosome is propably in charge of sex control (10, 11,12). The net morphology noticed in this research was as like as that found by (13), where all the autosomes had been metacentric. Secondary constriction was reported by (14) in pairs I, III and IV L. cluvia and L. sericata autosomes are well close pairing of somatic, an identified characteristic in Diptera, where the homologous chromosomes are line in neighborhood of one and the diploid complements show as haploid set (Agrawal et al. 2010). Nonetheless, XX of females and XY of males did not give like this s somatic pairing and line separately (15,16,17).



Fig(1) A-C-banded Mitotic chromosomes of Cochliomyia hominivorax



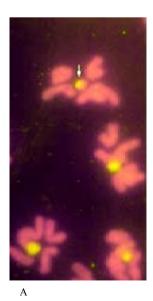
Fig(1) B- Mitotic chromosomes of Chrysomya bazziana

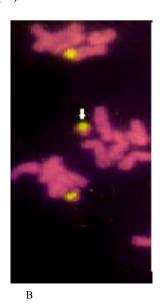
Chromosome pair	Length (µm)	Arm ratio	Relative	Designation
Chrysomya bazziana				
1	7.0	1.5	0.21	M
2	7.3	1.4	0.19	M
3	6.5	1.5	0.18	M
4	6.4	1.3	0.16	M
5	5.1	1.6	0.14	M
X	4.5	4.4	0.13	M
Y	2.7	1.2	0.06	M
Cochliomyia hominivorax				
1	7.4	1.4	0.20	M
2	6.7	1.5	0.18	M
3	6.7	1.2	0.17	M
4	5.8	1.5	0.15	M
5	5.3	1.2	0.13	M
X	6.4	1.2	0.15	M
Y	1.9	1.0	0.0	M

Table: Analysis of the somatic complements of *Chrysomya bazziana and Cochliomyia hominivorax* M: metacentric; St: sub telocentric. The relative length of Y was expressed as a function of the length of X.

The sex chromosomes contains NOR in both species according to in situ hybridization . Fig2(A and B). The NORs are positioned on the sex chromosomes in most species that studied on the sex chromosomes (18,19,20). *Lucilia cuprina* karyotype showed that the nucleolus has relatedness with X and Y secondary constrictions . (18)characterized a confirmed marker for rDNA in the sex chromosomes of *L. cuprina* and *C. bezziana*, as well as the regions had relatedness with secondary constrictions.

Cryptic or isomorphic species are present among blowflies that bring about to taxonomic problems seeing that the similarity of outer appearance of maggot a. Based on results got, rDNA could be taken in account as basic cytological sign for the comparison of karyotypes of phylogenetically related species and sibling species. These approaches can be used to participitate in the changes analysis of karyotype associated with the evolution process and to understand of taxonomic relations (20).





Fig(2) A- FISH in Cochliomyia hominivorax meiotic chromosomes.

Fig(2) B- FISH in Chrysomya bazziana

REFERENCES

- [1] Norris, K.R., and Murray, M.D(1964)Notes on the screw-worm fly Chrysomya (Dai ptera Calliphoridae) as a pest of cattle in New Guinea. CSIRO Division of EntomologyTechnical Paper No. 6.
- [2] Greenberg B 1973. Flies and Disease, Vol. 2, PrincetonUniversity Press, Princeton.
- [3] Spradbery, J.P., and Vanniasingham, J(1980) Incidence of the screwworm fly *Chrysomya bezziana*, at the Zoo Negara, Malaysia. Malays. Vet. J. 7: 28-32.
- [4]Greenberg B 1971. Flies and Disease, Vol. 1, Princeton University Press, Princeton.
- [5] Parise-Maltempi PP, Avancini RMP (2000) Cytogenetics of the neotropical flesh fly *Pattonella intermutans* (Diptera, Sarcophagidae). Genetics and Molecular Biology 23(3): 563–567. doi: 10.1590/S1415-47572000000300011.
- [6] Parise-Maltempi PP, Avancini RMP (2001) C-banding and FISH in chromosomes of the blow flies *Chrysomia megacephala* and *Chrysomia putoria* (Diptera, Calliphoridae). Memórias do Instituto Oswaldo Cruz 96(3): 371–377.
- [7] Parise-Maltempi PP, Avancini RMP (2007) Comparative cytogenetic study in Muscidae flies. Brazilian Journal of Biology 67(4): 945–950
- [8] Chirino MG, Rossi LF, Bressa MJ, Luaces JP, Merani MS (2014) Dipteran chromosomes: a simple method for obtaining high quality chromosomal preparations. Current Science 107(11): 1792–1794.
- [9] Levan A, Fredga K, Sanderg AA 1964. Nomenclature for centromeric position on chromosomes. *Hereditas57*: 201-220.
- [10] Üllerich FH (1973) Die genetiche Grundlage der Monogenie bei der Schmeibfliege *Chysomya rufifacies* (Calliphoridae, Diptera). *Mol Gen Genet125*: 157-172.
- [11] Üllerich FH(1976) Chromosomenverhältnisse, konstitutives hetrochromatin und geschlechtsbestimmung bei einigen Arten der Gattung *Chrysomya* (Calliphoridae, Diptera).
- [12] Bedo DG (1992) Nucleolar fragmentation in polytene trichogen cells of *Lucilia cuprina* and *Chrysomya bezziana* (Diptera: Calliphoridae). Genome 35(2): 283–293. doi: 10.1139/g92-
- [13] Boyes JW, Shewell GE (1975) Cytotaxonomy of Calliphoridae (Diptera). Genetica 45(4): 435–488. doi: 10.1007/bf01772870. 044
- [14] Azeredo-Espin AML, Pavan C (1983) Karyotypes and possible regions of origin of three species of Calliphoridae (Diptera) recently introduced in Brazil. Brazilian Journal of Genetics 6(4): 619–638.
- [15] Ullerich FH, Schöttke M (2006) Karyotypes, constitutive heterochromatin, and genomic DNA values in the blowfly genera *Chrysomya*, *Lucilia*, and *Protophormia* (Diptera: Calliphoridae). Genome 49(6): 584–597.
- [16] Agrawal UR, Bajpai N, Kurahashi H, Tewari RR (2010) Metaphase karyotypes of four species of Calliphoridae (Diptera). Chromosome Science 13: 49–52.
- [17] Holecová M, Rożek M, Maryańska-Nadachowska A, Jánošková V (2012) Karyotype of the bird blowfly, *Protocalliphora falcozi* Séguy, 1928 (Diptera: Calliphoridae). Folia Biologica 60(3-4): 129–133.
- [18] Bedo DG, Howells AJ (1987) Chromosomal localization of the white gene of *Lucilia cuprina* (Diptera; Calliphoridae) by *in situ* hybridization. Genome 29(1): 72–75. doi: 10.1139/g87-012.
- [19] Willhoeft U, Franz G (1996) Comparison of the mitotic karyotypes of *Ceratitis capitata*, *Ceratitis rosa*, and *Trirhithrum coffeae* (Diptera: Tephritidae) by C-banding and FISH. Genome 39(5): 884–889.
- [20] Willhoeft U (1997) Fluorescence *in situ* hybridization of ribosomal DNA to mitotic chromosomes of tsetse flies (Diptera: Glossinidae: Glossina). Chromosome Research 5(4): 262–267.

An open access journal of International Journal Foundation