

GENETIC POLYMORPHISM OF CYP2C19 IN A SAMPLE OF IRAQI POPULATION

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ABSTRACT

Background: CYP2C19 is responsible for the metabolism of a wide variety of medications this enzyme encoded by highly polymorphic gen. **Aim:** the aim of this study was to determine the frequencies of CYP2C19*1*2*3 and*17 in a sample of Iraqi population. **Methods:** This study was conduct in 221 Iraqi subjects. DNA was extracted from blood sample and detection of CYP2C19*1*2*3 and*17 was done by using TaqMan assay. **Results:** The frequencies for CYP2C19*1, CYP2C19*17, CYP2C19*2 and CYP2C19*3 were 65.1%, 19.5%, 15.2% and 0.2 respectively. Regarding phenotype the frequencies of EM, IM, UM and PM were 43.9 %, 27.1%, 27.1% and 1.9% respectively. **Conclusions:** The most common variant allele in this study was CYP2C19*17 and the EM was the most common phenotype

KEY WORDS

Cytochrome P450 - Iraqi - pharmacogenetics - polymorphism - SNP

INTRODUCTION

The CYP2Cs form an important subfamily of CYP enzymes that are responsible for metabolizing about 20% of most therapeutic drugs [1]. P450 2C19 is a member of four P450 2C monooxygenases that are encoded by a cluster of genes which began as gene duplication and divergence on chromosome 10. In human liver three of these P450s, 2C19, 2C9, and 2C8, are expressed. They significantly contribute to hepatic capacity to metabolize drugs [2]. CYP2C19 hydroxylates a wide variety of drugs like clopidogrel, lansoprazole, diazepam, barbiturates, nelfinavir, omeprazole, cyclophosphamide, and clonazepam [1]. CYP2C19 is extremely polymorphic and can cause difference in drug response. There are about 24 mutant allelic variants of CYP2C19 are known till now, of which CYP2C19*2, CYP2C19*3, and CYP2C19*17 are the most common [3].

Among *CYP2C19* defective alleles the most frequently founded one in human populations are *CYP2C19*2* (rs4244285). The *CYP2C19*2* is a defective allele that carries a 681G<A single base substitute in exon 5

causing an alternate splice site consensus and by this it eliminates the catalytic activity on all substrates [4]. The other important defective allele is CYP2C19*3 (rs4986893), this has an adenine substituted for a guanine at base 636 (636G>A) in exon 4, forming a premature stop codon [4]. The CYP2C19*17 allele (c.-806C>T; rs12248560) on the contrary results in increased activity as a result of enhanced transcription [5]. Several researches have investigated the frequency of different CYP2C19 alleles worldwide. The allele frequency of CYP2C9*2 was reported to be about 50% in Asians, 19% in American populations, 34% in Africans and 18% in Caucasians [6-9]. The allele frequency of CYP2C9*3 among the Asian, African and Caucasian populations was <7 %, <1 % and 1 %, respectively [10] The CYP2C19*17 genotype was found in 22,0 % of the Norse (Pedersen et al 2010), 25,7 % of the Germans [11], 20,0 % of the Swedes [12], 4,0 % of the Chinese [5], 0,3 % of the Koreans [13], 1,3 % of the Japanese [12], 25.7 % of Saudi Individuals [14].

CYP2C19 enzyme activity are considered to be normal when two *CYP2C19*1* alleles are present

54



(CYP2C19*1/*1); CYP2C19 Intermediate Metabolizer (IM) phenotype is expected by the existence of one CYP2C19 allele with normal function and one CYP2C19 allele with decreased function or one CYP2C19 allele with increased function and one CYP2C19 allele with decreased function (CYP2C19*1/*2, *1/*3, *2/*17, *3/*17); CYP2C19 Poor Metabolizer (PM) phenotype is expected when two CYP2C19 non-functional alleles CYP2C19*2 or CYP2C19*3 are present (CYP2C19*2/*2, *2/*3, *3/*3). In contrast presence of Heterozygosity or homozygosity for the increased function CYP2C19*17 allele is associated with CYP2C19 increased activity that will lead to an ultra-rapid metabolizer phenotype (UM) [15].

MATERIAL AND METHODS

Subject

Two hundred and twenty one of unrelated Iraqi nationality and Arab ethnicity (127 male and 94 female, aged 19 to 85 years) were enrolled in this study. Ethical approval of this study obtained from the Research and Ethics Committee of Medical College / Al-Nahrain University. All subjects included in this study informed about the test that will done in this study.

CYP 2C19 genotyping

For genetic study 2.5 ml of the blood was drawn from each subject and transferred in to EDTA containing tube. DNA was extracted by using a kit provided by promega (ReliaPrep[™] Blood gDNA Miniprep System). DNA extraction was done according to manufacturer's instructions. CYP2C19 *2,*3 and *17 were determined by TaqMan assay using STRATAGENE-MX3005P.

PCR amplification for all three SNP was done in 20 μ L reactions with about 80ng of template DNA, 1X KAPA

PROBE FAST qPCR Master Mix Universal, 600nm each primer, 400nm each probe, and deionized water. The thermal condition of the reaction began with denaturation at 95° C for 10 min, followed by 40 cycles of denaturation at 95° C for 30 sec, annealing, and extension at 60° C for 1 min. at the end of amplification results were obtained from the software supplied with STRATAGENE-MX3005P. As a quality control of the genotyping, 20 per cent of the total sample genotyped was sequenced.

RESULTS

The frequency of CYP2C19 alleles, genotype and phenotype in all subject enrolled in this study are shown in table 3-12. The most frequent allele was CYP2C19*1(288/441, 65%). CYP2C19*1*1 was the most frequent genotype (97/221, 44%). The most frequent variant allele was CYP2C19*17 (86/441, 19.5%). CYP2C19*17 where classified as 51 (23.1%) for CYP2C19*1*17, heterozygous 17 (7.7%) heterozygous for CYP2C19*2*17 and 9 (4.1%) homozygous for the CYP2C19*17*17. The second most common variant allele in this study was CYP2C19*2 (67/442, 15.2%), these allele are distributed as 43(19.5%) heterozygous for CYP2C19*1*2, 17 (7.7%) heterozygous for CYP2C19*17*2 and 3 (1.4%) homozygous for CYP2C19*2*2. There was only on CYP2C19*3 allele in this study which was heterozygous CYP2C19*2*3. for Regarding phenotype the frequencies of EM, IM, UM and PM were 43.9 %, 27.1%, 27.1% and 1.9% respectively. Based on Hardy-Weinberg equilibrium there was no statistical difference in the actual and expected frequency distribution (P>0.05)

55



Int J Pharm Biol Sci.

CYP2C19	Allele	Actual	ual 95% Cl		Expected by Hardy-Weinberg law	
		Number	Frequency		Number	Frequency
Alleles	*1	288	0.652	0.607-0.696	N/A	N/A
	*2	67	0.152	0.118-0.185	N/A	N/A
	*3	1	0.002	0.000-0.007	N/A	N/A
	*17	86	0.195	0.158-0.203	N/A	N/A
	Total	442	1.000	N/A	N/A	N/A
Phenotypes	Genotype					
EM	*1*1	97	0.439	0.373-0.504	93.828	0.425
	*1*17	51	0.231	0.175-0.286	56.036	0.254
	*17*17	9	0.041	0.015-0.067	8.367	0.038
	*1*2	43	0.195	0.142-0.247	43.656	0.198
	*2*17	17	0.077	0.042-112	13.036	0.059
	*1*3	0	0.000	N/A	0.652	0.003
	*3*17	0	0.000	N/A	0.195	0.001
	*2*2	3	0.014	0.000-0.029	5.078	0.023
	*2*3	1	0.005	0.000-0.013	0.152	0.001
	*3*3	0	0.000	N/A	0.001	0.000
	Total	221	1.000		221.000	1.000

Table 1: Frequencies of CYP2C19 alleles and genotypes in Iraqi population sample (n=221)

DISCUSSION

This study is the first to characterized the phenotype and genotype of CYP 2C19 in Iraqi population. The genetic polymorphism of CYP2C19 has been shown to have the most striking interethnic variation of a CYP so far. The PM frequency ranges from 2 to 7% in Caucasians, 14-25% in Asians, and 60% in the Vanuatu [16,17]. In this study the CYP2C19*2 frequency was 15.1%. This frequency was similar to that of Saudi Arabian 15 % [18] and higher than those reported in Caucasians (14.7%), Egyptian (11.0%) [19], while it was lower than Asians (30%) as shown in table 2. Moreover, this study revealed that the CYP2C19*3 frequency was 0.2% (only one subject). However there is limited numbers of studies have reported CYP2C19*3 in populations other than the Asians. CYP2C19*3 was reported in the Egyptian population 0.2% [19], and Swedish population 0.3% [20], 0.7% [21]. CYP2C19*3 has been considered as an Asian mutation allele and after genotyping for CYP2C19*2 it was responsible for the remaining alleles in Asian PMs [19].

Regarding CYP2C19*17 we found that the frequency of CYP2C19*17 was 19.4. The allele frequency of CYP2C19*17 in Iraqi population was similar to that found in African Americans 21% [22], Danish 20.1% [23] and Greece 19.6 [24]. While it was slightly lower

than Saudi Arabia 25% [14], German 25.5% [11] and Polish 27.2% [25]Lower frequencies of CYP2C19*17 were reported for in East and South Asian groups (Chinese, Japanese, and Koreans), the CYP2C19*17 allele frequencies (range: 1.2% -1.5%) [26, 27].

Based on the genotype results, the classification of patients was made according to their metabolizer phenotype. Phenotype classifications divided patients into those who were extensive (normal) metabolizers, intermediate metabolizers, poor metabolizers and rapid metabolizers.

Our result revealed that the EM phenotype in 221 Iraqi subjects was 43.9% while the frequency of IM in our study was 27.15%, in addition our study demonstrate that the frequency of PM and UM were 1.8% and 27.15% respectively. Similar results were obtained by Leena et al and Goldstein et al in Saudi Arabian population [14, 18].

In addition the result of this study was similar to that obtained in pan-ethnic population. The metabolic rate in pan- ethnic population include the following: 41% of populations were extensive metabolizer, 27.57% as intermediate metabolizer, 27.8% as ultrarapid metabolizer and 3.46 as poor metabolizer [28].

In Asian populations the frequencies of poor and intermediate metabolizer were higher than that observed in this study while ultrarapid metabolizer was lower. In Thai population the frequencies of



Int J Pharm Biol Sci.

intermediate, poor and ultrarapid metabolizer were 41.95%, 13.03% and 4.3% respectively [29]. In Korean population the frequency of poor and intermediate metabolizer were 12%, 42% respectively [30]. This can be explain by Lower frequencies of CYP2C19*17 and higher frequencies of CYP2C19*3 in these population [12, 26, 27].

CONCLUSION

In term of genotyping, the most frequent allele in a sample of Iraqi population was CYP2C19*1, while the most frequent variant allele was CYP2C19*17. In term of phenotype, Extensive metabolizer was the most frequent phenotype of CYP2C19, while Ultrarapid and intermediate metabolizer have the same frequency

Table 2: Ethnic variation of CYP2C19 (*1, *2, *3, and *17) in the present study and pub

population	Number	Alleles frequency of CYP2C19				Reference
		*1	*2	*3	*17	
Iraqi population	221	65.1	15.2	0.2	19.4	Our study
Saudi Arabians	97	85	15	0.00	NA	[18]
Saudi Arabians	201	62.8	11.5	NA	25.7	[14]
Egyptians	494	88	12	6	NA	[19]
Jordania	78	84	16	0.0	NA	[31]
Lebanese	161	83.6	13.4	3	NA	[32]
Tunisian region	100	88.5	11.5	NA	NA	[33]
Iranian:Tehran	200	86	14	0	NA	[34]
Iranian:Turkman	140	56.4	23.5	20	NA	[35]
Japanese	217	61.2	27.4	10.8	NA	[36]
Japanese	265	70.8	27.9	NA	1.3	[12]
Chinese	121	49.5	45.5	4.5	NA	[37]
Chinese	384	73.9	24.9	NA	1.2	[26]
Koreans	103	67.4	20.9	11.7	NA	[30]
Thai	774	68	29	3	NA	[38]
Thais	1051	63	27	6	4	[29]
Burmese	127	66	30	4	NA	[38]
Malaysians	142	66	28	6	NA	[39]
German	237	59.3	15.2	NA	25.5	[11]
Danish	276	64.9	15.0	NA	20.1	[23]
Polish	125	61.2	11.6	NA	27.2	[25]
Greece	283	79.4	NA	NA	19.6	[24]
Swedish	310	81.2	18.8	0	NA	[21]
Italians	360	88.9	11.1	0	NA	[40]
Croatians	200	85	15	0	NA	[41]
Belgian	121	90.9	9.1	0	NA	[42]
Islands	5538	22.3	63.3	14.4	NA	[43]
Ethiopians	114	83	14	3	NA	[44]
Tansanians	251	81	18	1	NA	[45]
Russians	290	88.3	11.4	0.3	NA	[46]
African-American	114	79	NA	NA	21.0	[22]
African Americans	517	81	19	0	NA	[47]
Americans	100	80	11	0	9	[48]

NA: not analyzed



REFERENCES

- [1]Goldstein JA. Clinical relevance of genetic polymorphisms in the human CYP2C subfamily. Br J Clin Pharmacol, 52: 349-355, (2001)
- [2]Zanger, U. M., Turpeinen, M., Klein, K., and Schwab, M.
 Functional pharmacogenetics/genomics of human cytochromes P450 involved in drug biotransformation.
 Anal. Bioanal. Chem, 392: 1093–1108, (2008)
- [3]Sibbing D, Stegherr J, Latz W, et al. Cytochrome P450 2C19 loss-of-function polymorphism and stent thrombosis following percutaneous coronary intervention. Eur Heart J, 30(8): 916–922, (2009)
- [4]De Morais SM, Wilkinson GR, Blaisdell J, et al. Identification of a new genetic defect responsible for the polymorphism of (S)-mephenytoin metabolism in Japanese. Mol Pharmacol, 46:594-602, (1994)
- [5]Sim SC, Risinger C, Dahl ML, et al. A common novel CYP2C19 gene variant causes ultrarapid drug metabolism relevant for the drug response to proton pump inhibitors and antidepressants. Clin Pharmacol Ther,79:103–113, (2006)
- [6]Luo HR, Poland RE, Lin KM, et al. Genetic polymorphism of cytochrome P450 2C19 in Mexican Americans: a cross-ethnic comparative study. Clin Pharmacol Ther, 80: 33-40, (2006).
- [7]Mega JL, Close SL, Wiviott SD, et al. Cytochrome p-450 polymorphisms and response to clopidogrel. New England Journal of Medicine, 360(4):354-362, (2009)
- [8]Giusti B, Gori AM, Marcucci R, et al. Relation of cytochrome P450 2C19 loss-of-function polymorphism to occurrence of drugeluting coronary stent thrombosis. Am J Cardiol, 103: 806-811, (2009).
- [9]Bonello L, Armero S, Ait Mokhtar O, et al. Clopidogrel loading dose adjustment according to platelet reactivity monitoring in patients carrying the 2C19*2 loss of function polymorphism. J Am Coll Cardiol, 56: 1630-1636, 2010.
- [10]Collet JP, Hulot JS, Pena A, et al. Cytochrome P450 2C19 polymorphism in young patients treated with clopidogrel after myocardial infarction: a cohort study. Lancet, 373:309 –17, (2009)
- [11]Geisler T, Schaeffeler E, Dippon J, et al (2008). CYP2C19 and nongenetic factors predict poor responsiveness to clopidogrel loading dose after coronary stent implantation. Pharmacogenomics; 9: 1251-1259.
- [12]Sugimoto K, Lino T, Yamazalci H, et al. Limited frequency of the CYP2C19*17 allele and its minor role in a Japanese population. Br J Clin Pharmacol, 65: 437-439, (2008)
- [13]Ramsjö M, Aklillu E, Bohman L, et al. CYP2C19 activity comparison between Swedes and Koreans: effect of

genotype, sex, oral contraceptive use, and smoking. Eur J Clin Pharmacol, 66: 871-877, (2010)

- [14]Leena H Saeed and Ahmed Y Mayet. Genotype-Phenotype Analysis of CYP2C19 in Healthy Saudi Individuals and its Potential Clinical Implication in Drug Therapy.International Journal of Medical Sciences, 10(11):1497-1502, (2013).
- [15]Scott S A, Sangkuhl K, Stein CM, J et al. Clinical Pharmacogenetics Implementation Consortium Guidelines for CYP2C19 Genotype and Clopidogrel Therapy: 2013 Update. Clinical pharmacology & Therapeutics, 94 (3): 317-23, (2013).
- [16]Bertilsson L, Dahl ML, Ingelman-Sundberg M,et al. Interindividual and interethnic differences in polymorphic drug oxidation-implications for drug therapy with focus on psychoactive drugs. In Advances in Drug Metabolism in Man, eds Pacifici GM, Fracchia GN. DGXII-E-4eur1549 EN, EC. Office for Official Publications of the European communities, Luxembourg, 1995; 85–136, (1995)
- [17]Kaneko A, Berqvist Y, Taleo G, et al. Proguanil disposition and toxicity in malaria patients from Vanuatu with high frequencies of CYP2C19 mutations. Pharmacogenetics, 9: 317–326, (1999)
- [18]Goldstein JA, Ishizaki T, Chiba K, et al. Frequencies of the defective CYP2C19 alleles responsible for the meph enytoinpoor metabolizer phenotype in various Oriental, Caucasian, Saudi Arabian and American black populations. Pharmacogenetics, 7: 59-64, (1997)
- [19]Hamdy SI, Hiratsuka M, Narahara K, et al. Allele and genotype frequencies of polymorphic cytochromes P450 (CYP2C9, CYP2C19, CYP2E1) and dihydropyrimidine dehydrogenase (DPYD) in the Egyptian population. Br J Clin Pharmacol, 53:596-603, (2002)
- [20]Chang M, Dahl ML, Tybring G, et al. Use of omeprazole as a probe drug for CYP2C19 phenotype in Swedish Caucasians: comparison with S-mephenytoin hydroxylation phenotype and CYP2C19 genotype. Pharmacogenetics, 5: 358–363, (1995)
- [21]Yamada H, Dahl M-L, Lannfelt L, et al. CYP2D6 and CYP2C19 genotypes in an elderly Swedish population. Eur J Clin Pharmacol, 54: 479–481, (1998)
- [22]Kearns GL, Leeder JS, Gaedigk A. Impact of the CYP2C19*17 allele on the pharmacokinetics of omeprazole and pantoprazole in children: evidence for a differential effect. Drug Metab Dispos., 38: 894-897, (2010)
- [23]Pedersen RS, Brasch-Andersen C, Sim SC, Bergmann TK, Hailing J, Petersen MS et al. Linkage disequilibrium between the CYP2C19*17 allele and wildtype CYP2C8 and CYP2C9 alleles: identification of CYP2C haplotypes in healthy Nordic populations. Eur J Clin Pharmacol, 66: 1199-1205, (2010).



- [24]Ragia G, Arvanitidis KI, Tavridou A, et al. Need for reassessment of reported CYP2C19 allele frequencies in various populations in view of CYP2C19*17 discovery: the case of Greece. Pharmacogenomics,10: 43-49, (2009)
- [25]Kurzawski M, Gawronska-Szldarz B, Wrzesniewska J,et al. Effect of CYP2C19*17 gene variant on Helicobacter pylori eradication in peptic ulcer patients. Eur J Clin Pharmacol, 62: 877-880, (2006)
- [26]Chen L, Qin S, Xie J, et al. Genetic polymorphism analysis of CYP2C19 in Chinese Han populations from different geographic areas of mainland China. Pharmacogenomics, 9: 691-702, (2008).
- [27]Kim KA, Song WK, Kim KR et al. Assessment of CYP2C19 genetic polymorphisms in a Korean population using a simultaneous multiplex pyrosequencing method to simultaneously detect the CYP2C19*2, CYP2C19*3, and CYP2C19*17 alleles. J Clin Pharm Ther, 35: 697-703, (2010)
- [28]Charles M. Strom, Dana Goos, et al. Testing for variants in CYP2C19: population frequencies and testing experience in a clinical laboratory. Genetics in medicine, 14 (1): 95-100, (2012)
- [29]Chonlaphat Sukasem, Ramaimon Tunthong, Montri Chamnanphon, et al. CYP2C19 polymorphisms in the Thai population and the clinical response to clopidogrel in patients with atherothrombotic-risk factors. Pharmacogenomics and Personalized Medicine, 6:85– 91, (2013)
- [30]Roh HK, Dahl ML, Tybring G, et al. CYP2C19 genotype and phenotype determined by omeprazole in a Korean population. Pharmacogenetics, 6(6):547-551, (1996)
- [31]Zalloum I, Hakooz N, Arafat et al.Genetic polymorphism of CYP2C19 in a Jordanian population: influence of allele frequencies of CYP2C19*1 and CYP2C19*2 on the pharmacokinetic profile of lansoprazole. Mol Biol Rep.,39(4):4195-200, (2012)
- [32]Djaffar Jureidini I, Chamseddine N, Keleshian S, et al. Prevalence of CYP2C19 polymorphisms in the Lebanese population. Mol Biol Rep, 38(8):5449-52, (2011)
- [33]Leila Abid, Lobna Laroussi1, Amine Bahloul, et al. Impact of cytochrome P450 2C19*2 polymorphism on the clinical cardiovascular events after stent implantation in patients receiving clopidogrel of a southern Tunisian region. World Journal of Cardiovascular Diseases, 3: 4-10, (2013)
- [34]Zand N, Tajik N, Moghaddam AS, et al. Genetic polymorphisms of cytochrome P450 enzymes 2C9 and 2C19 in a healthy Iranian population. Clin Exp Pharmacol Physiol, 34(1-2):102-105, (2007)
- [35]Robabeh Ghiyas Tabari, Abdoljalal Marjani, Ogholdondy Agh Ataby, et al. Genetic Polymorphism of Cytochrome

p450 (2C19) Enzyme in Iranian Turkman Ethnic Group. Oman Medical Journal, 28(4):237-244, (2013)

- [36]Takakubo F, Kuwano A, and Kondo I. Evidence that poor metabolizers of (S)- mephenytoin could be identified by haplotypes of CYP2C19 in Japanese. Pharmacogenetics, 6(3):265-267, (1996)
- [37]Yamada S, Onda M, Kato S, et al. Genetic differences in CYP2C19 single nucleotide polymorphisms among four Asian populations. J Gastroenterol, 36(10):669-672, (2001)
- [38]Tassaneeyakul W, Mahatthanatrakul W, Niwatananun K, et al. CYP2C19 genetic polymorphism in Thai, Burmese and Karen populations. Drug Metab Pharmacokinet, 21(4):286-290, (2006)
- [39]Pang YS, Wong LP, Lee TC, et al. Genetic polymorphism of cytochrome P450 2C19 in healthy Malaysian subjects. Br J Clin Pharmacol, 58(3):332-335,(2004)
- [40]Scordo MG, Caputi AP, D'Arrigo C, et al. Allele and genotype frequencies of CYP2C9, CYP2C19 and CYP2D6 in an Italian population. Pharmacol Res, 50(2):195-200, (2004)
- [41]Boina N, Graniæ P, Laliæ Z, et al. Genetic polymorphisms of cytochromes P450: CYP2C9, CYP2C19, and CYP2D6 in Croatian Population. Roatian Med J, 44(4):425-428, (2003)
- [42]Allabi AC, Gala JL, Desager JP, et al. Genetic polymorphisms of CYP2C9 and CYP2C19 in the Beninese and Belgian populations. Br J Clin Pharmacol, 56(6):653-657, (2003)
- [43]Kaneko A , Lum JK, Yaviong L, et al. High and variable frequencies of CYP2C19 mutations: medical consequences of poor drug metabolism in Vanuatu and other Pacific islands. Pharmacogenetics, 9(5):581-590, (1999).
- [44]Persson I, Aklillu E, Rodrigues F, et al. S-mephenytoin hydroxylation phenotype and CYP2C19 genotype among Ethiopians. Pharmacogenetics, 6(6):521-526, (1996)
- [45]Herrlin K, Massele AY, Jande M, et al. Bantu Tanzanians have a decreased capacity to metabolize omeprazole and mephenytoin in relation to their CYP2C19 genotype. Clin Pharmacol Ther ,64(4):391-401, (1998).
- [46]Gaikovitch EA, Cascorbi I, Mrozikiewicz PM, et al. Polymorphisms of drug-metabolizing enzymes CYP2C9, CYP2C19, CYP2D6, CYP1A1, NAT2 and of P-glycoprotein in a Russian population. Eur J Clin Pharmacol, 59(4):303-312, (2003)
- [47]Xie HG, Kim RB, Stein CM, et al. Genetic polymorphism of (S)-mephenytoin 4'-hydroxylation in populations of African descent. Br J Clin Pharmacol, 48(3):402-408, (1999)
- [48]Oestreich J. H., Best L. G. and Dobesh P. Prevalence of CYP2C19 variant alleles and pharmacodinamic variability

Hussein Ali Sahib* et al

59

Int J Pharm Biol Sci.



of aspirin and clopidogrel in Native Americans. Am. Heart J., 167, 413–418,(2014).

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Hussein Ali Sahib* et al

60

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